# organic papers

Acta Crystallographica Section E Structure Reports Online

ISSN 1600-5368

# Muhammad Sajid,<sup>a</sup> Shazia Anjum,<sup>a</sup> M. Iqbal Choudhary,<sup>a</sup> Atta-ur-Rahman,<sup>a</sup> Masood Parvez<sup>b</sup>\* and letidal Eltahir Mohamed<sup>c</sup>

<sup>a</sup>International Centre for Chemical Sciences, HEJ Research Institute of Chemistry, University of Karachi, Karachi 75270, Pakistan, <sup>b</sup>Department of Chemistry, 2500 University drive NW, Calgary, Alberta, Canada T2N 1N4, and <sup>c</sup>Botany Department, University of Khartoum, Khartoum, PO Box 321, PC 11115, Sudan

Correspondence e-mail: parvez@ucalgary.ca

#### Key indicators

Single-crystal X-ray study T = 293 K Mean  $\sigma$ (C–C) = 0.006 Å R factor = 0.067 wR factor = 0.120 Data-to-parameter ratio = 16.1

For details of how these key indicators were automatically derived from the article, see http://journals.iucr.org/e.

# Absolute configuration of (*R*)-synephrine hydrochloride

The title compound, 2-hydroxy-2-(4-hydroxyphenyl)-*N*-methylethanaminium chloride,  $C_9H_{14}NO_2^+ \cdot Cl^-$ , has been isolated for the first time from the genus *Harrisonia*. Its structure is stabilized by extensive intra- and intermolecular hydrogen bonds of type  $O-H\cdots O$ ,  $N-H\cdots O$ ,  $N-H\cdots Cl$  and  $C-H\cdots Cl$ , resulting in a three-dimensional network. Additional weak  $\pi$ - $\pi$  stacking interactions between adjacent molecules further stabilize the crystal structure.

## Comment

Harrisonia abyssinica Oliv. (Simaroubaceae) is widely used in various African folk remedies. Its roots and stem bark are used throughout Africa in the treatment of gonorrhea, dysentery, skin diseases, tuberculosis, bilharzia infections and as an ascaricide (Balde et al., 1989; Chhabra et al., 1984; Sofowora, 1982). The plant has also been reported to show antifeedant activity against the most common East African crop pests, the monophageous Sophoptera exempta (African army worm) and S. eridania (Southern army worm), and antibacterial activity against Gram-positive microorganisms, as well as cytotoxic and plant-growth inhibitory activities (Kubo et al., 1976). The butanolic extract of the fruit of Harrisonia abyssinica was phytochemically investigated to give the compound synepherine hydrochloride, (I). The natural occurrence of synepherine has been reported in various genera, such as Citrus, Coryphantha, Ficus and Haloxylon (Stewart & Wheaton, 1964; Stewart et al., 1964; Ranieri & Mclaughlin, 1976; Smith, 1977). However, this is the first time that synepherine hydrochloride has been isolated from the genus Harrisonia.



Synepherine is a sympathomimetic agent showing vasoconstrictor, hypertensive and bronchial muscle relaxant properties (Chen *et al.*, 1980; Stewart *et al.*, 1964). The absolute stereochemistry of synepherine was determined by X-ray diffraction studies of its synthetic salt as the (-)-3-bromocamphor-8-sulfonate (Midgley *et al.*, 1989). In this paper, we report the absolute stereochemistry of naturally occurring (*R*)-synepherine hydrochloride, (I).

The bond lengths and bond angles in (I) (Fig. 1) are within normal ranges (Allen *et al.*, 1987). Atoms C7 and O1 are

Printed in Great Britain - all rights reserved

© 2005 International Union of Crystallography

Received 5 July 2005 Accepted 7 July 2005 Online 16 July 2005



Figure 1 An ORTEPII (Johnson, 1976) drawing of (I), showing 50% probability displacement ellipsoids and the atomic numbering scheme.

essentially in the plane of the benzene ring. The hydroxyl group at C7 is twisted away from the benzene ring, the O2-C7-C1-C2 torsion angle being 131.5 (4)°. The side chain comprising atoms C1, C7, C8, N1 and C9 is fully extended and its mean plane is inclined at an angle of  $71.9(2)^{\circ}$  with respect to the mean plane of the benzene ring.

The anions and the cations in (I) are linked together to form a three-dimensional network of hydrogen bonds (Fig. 2). In addition to N-H···O and C-H···Cl intramolecular hydrogen bonds, there are four intermolecular hydrogen bonds of the types  $O-H\cdots Cl$ ,  $N-H\cdots Cl$  and  $O-H\cdots O$ , which stabilize the crystal structure of (I); details of the hydrogen-bonding geometries are given in Table 1. The stability of the structure is also achieved by  $\pi - \pi$  stacking interactions between benzene rings from adjacent molecules. The mean planes of these rings are 3.150 (3) Å apart, indicating weak  $\pi - \pi$  interactions, with the distance between the ring centroids being 4.785 (3) Å.

# **Experimental**

Powdered fruits of Harrisonia abyssinica (2.4 kg) were soaked and extracted with EtOH (101) for one week. The combined EtOH extracts were dried under vacuum to furnish a brown gum (150 g). Subsequent extraction of this EtOH extract afforded a hexanesoluble fraction (10 g), an ethyl acetate-soluble fraction (40 g) and a butanol-soluble fraction (5.5 g). The butanolic extract (5.5 g) was passed through a polyamine column and ten fractions were obtained. Fractions 3–5 were found to be similar by thin-layer chromatography and were combined (2.1 g). These combined fractions were again subjected to flash chromatography, using dichloromethane and methanol as the eluting solvents, and 14 fractions were obtained. Fractions 7-10 showed the formation of some needles which could not be separated by washing, so they were combined to give the subfraction HBR (200 mg). Sub-fraction HBR was again subjected to flash chromatography, using dichloromethane and methanol as the eluting solvents, to give compound (I) in a semi-purified form. This was further purified by recrystallization from methanol and dichloromethane (1:1) (10 mg, m.p. 508 K).



Figure 2

A packing diagram of (I). Hydrogen bonds are represented by broken lines.

Mo  $K\alpha$  radiation

reflections

 $\theta=1.7{-}25.0^\circ$ 

 $\mu = 0.32 \text{ mm}^{-1}$ 

T = 293 (2) K

 $R_{\rm int}=0.058$ 

 $\theta_{\rm max} = 25.0^{\circ}$ 

 $h = -8 \rightarrow 7$ 

 $k = -8 \rightarrow 8$ 

 $l = -27 \rightarrow 25$ 

Needle, colourless

 $0.58 \times 0.06 \times 0.05 \text{ mm}$ 

1928 independent reflections

1400 reflections with  $I > 2\sigma(I)$ 

Cell parameters from 6750

Crystal data

C<sub>9</sub>H<sub>14</sub>NO<sub>2</sub><sup>+</sup>·Cl<sup>-</sup>  $M_r = 203.66$ Orthorhombic,  $P2_12_12_1$ a = 6.7480 (13) Åb = 6.9559 (13) Å c = 23.450 (4) Å V = 1100.7 (4) Å<sup>3</sup> Z = 4 $D_x = 1.229 \text{ Mg m}^{-3}$ 

#### Data collection

Siemens SMART CCD areadetector diffractometer  $\omega$  scans Absorption correction: multi-scan (SADABS; Sheldrick, 1996)  $T_{\min} = 0.837, \ T_{\max} = 0.984$ 5679 measured reflections

#### Refinement

Refinement on $F^2$	$w = 1/[\sigma^2(F_o^2) + (0.0489P)^2$
$R[F^2 > 2\sigma(F^2)] = 0.067$	+ 0.1384P]
$wR(F^2) = 0.120$	where $P = (F_0^2 + 2F_c^2)/3$
S = 1.02	$(\Delta/\sigma)_{\rm max} < 0.001$
1928 reflections	$\Delta \rho_{\rm max} = 0.30 \text{ e } \text{\AA}^{-3}$
120 parameters	$\Delta \rho_{\rm min} = -0.16 \text{ e } \text{\AA}^{-3}$
H-atom parameters constrained	Absolute structure: Flack (1983),
	with 764 Friedel pairs
	Flack parameter: 0.07 (14)

Table 1 Hydrogen-bond geometry (Å, °).

$D - H \cdots A$	D-H	$H \cdot \cdot \cdot A$	$D \cdots A$	$D - \mathbf{H} \cdot \cdot \cdot A$
$D1 - H1 \cdots Cl1^{i}$ $N1 - H1 A \cdots Cl1^{ii}$	0.82	2.23	3.037 (3) 3.091 (4)	168 163
$N1 - H1B \cdot \cdot \cdot O2^{iii}$	0.90	2.42	2.769 (4)	103
$N1 - H1B \cdots Cl1^{v}$ $O2 - H2 \cdots O1^{v}$	0.90 0.82	2.31 2.01	3.161 (3) 2.825 (5)	157 171
$C8-H8A\cdots Cl1$	0.97	2.82	3.603 (4)	138
Symmetry codes: (i)	$x - \frac{1}{2}, -y + \frac{1}{2}$	, -z; (ii) $x$	-1, y, z; (iii)	x, y, z; (iv)

 $-x + 1, y - \frac{1}{2}, -z + \frac{1}{2};$  (v)  $x - \frac{1}{2}, -y - \frac{1}{2}, -z.$ 

All H atoms were positioned geometrically and were allowed to ride on their parent atoms, with aromatic C-H = 0.93, methyl C-H =0.96 Å, N-H = 0.90 Å and O-H = 0.82 Å, and with  $U_{iso}(H) =$   $1.5U_{eq}(C)$  for methyl H and  $1.2U_{eq}(parent)$  for other H atoms. The chirality of the structure was determined from the Flack (1983) parameter, the value for the inverted structure being 0.93 (14).

Data collection: *SMART* (Siemens, 1996); cell refinement: *SAINT* (Siemens, 1996); data reduction: *SAINT*; program(s) used to solve structure: *SHELXTL* (Sheldrick, 1997); program(s) used to refine structure: *SHELXTL*; molecular graphics: *SHELXTL*; software used to prepare material for publication: *SHELXTL*, *PARST95* (Nardelli, 1995) and *PLATON* (Spek, 2003).

SA thanks the Higher Education Commission, Government of Pakistan, for research grant R&D/Acad/03/1064. MP thanks the International Centre for Chemical Sciences, HEJ Research Institute of Chemistry, University of Karachi, Pakistan, for sponsoring his visit to Pakistan.

### References

Allen, F. H., Kennard, O., Watson, D. G., Brammer, L., Orpen, A. G. & Taylor, R. (1987). J. Chem. Soc. Perkin Trans. 2, pp. S1–19.

- Balde, A. M., Van Marck, E., Kestens, L., Gigase, P. L. & Vlietnick, A. J. (1989). Planta Med. 55, 41–43.
- Chen, X., Huang, Q. & Zhou, T. (1980). Acta Pharm. Sin. 15, 71-73.
- Chhabra, S. C., Uiso, F. C. & Mshiu, E. N. (1984). J. Ethnopharmacol. 11, 157– 158.
- Flack, H. D. (1983). Acta Cryst. A39, 876-881.
- Johnson, C. K. (1976). ORTEP II. Report ORNL-5138. Oak Ridge National Laboratory, Tennessee, USA.
- Kubo, I., Tanis, S. P., Lee, Y. M., Miura, I., Nakanishi, K. & Chapaya, A. (1976). *Heterocycles*, 5, 485–487.
- Midgley, J. M., Thonoor, C. M., Darke, A. F., William, C. M., Koziol, A. E. & Palenik, G. J. (1989). J. Chem. Soc. Perkin Trans. 2, pp. 963–969.
- Nardelli, M. (1995). J. Appl. Cryst. 28, 659.
- Ranieri, R. L. & Mclaughlin, J. R. (1976). J. Org. Chem. 41, 319-323.
- Sheldrick, G. M. (1996). SADABS. University of Göttingen, Germany.
- Sheldrick, G. M. (1997). SHELXTL. Version 5.10. Bruker AXS Inc., Madison, Wisconsin, USA.
- Siemens (1996). SMART and SAINT. Siemens Analytical X-Ray Instruments Inc., Madison, Wisconsin, USA.
- Smith, T. A. (1977). Phytochemistry, 16, 9-18.
- Sofowora, A. (1982). Medicinal Plants and Traditional Medicine in Africa, edited by M. Kaumare, pp. 223–223. Chichester: Wiley.
- Spek, A. L. (2003). J. Appl. Cryst. 36, 7-13.
- Stewart, I., Newhall, W. F. & Edwards, G. J. (1964). J. Biol. Chem. 239, 939–932. Stewart, I. & Wheaton, T. A. (1964). Science, 145, 60–61.